

STUDIES ON BIOCHEMICAL CHANGES DURING LARVA TO PUPA TRANSFORMATION IN ANTHEREAEMYLITTADRURY (DABA TV)

Lakshmi Marepally

*Post - Doctoral Fellow-UGC, Department of Zoology,
Kakatiya University, Warangal, Telangana, (India)*

ABSTRACT

*The present study has been carried out to analyze the biochemical variations in *Antheraea mylittadrury* (Daba TV) during larval to pupal transformation. In this study, various biochemical assays have been performed in haemolymph and fat body of silkworms during 1st to 6th day of 5th instar larvae, spinning day 1, spinning day 2 and pre pupal stages. The results revealed that during larval to pupal transformation a drastic variation in proteins, amino acids, haemocytes and acid phosphatase activity has been identified. An increase in protein content of both haemolymph and fat body was noticed upto 6th day of fifth instar larvae but a gradual reduction in protein content was observed during first day of spinning, second day of spinning and also in prepupal stage. A reduction in amino acid levels of both haemolymph and fat body was recorded till 6th day of fifth instar larvae whereas an increase is noticed in spinning day 1, 2 and prepupa. Present results also shows an increase in haemocyte count till 6th day of fifth instar and a later decrease in spinning day 1, 6 and pupal stages. Significant increase in the activity of Acid phosphatase was observed in the fifth instar from 1st to 6th days and also during spinning day 1, spinning day 2 and prepupal stages.*

Keywords: *Haemolymph, Fatbody, Proteins, Aminoacids, Haemocytes, Acid phosphatase, Transformation.*

I. INTRODUCTION

Antheraea mylitta Drury a lepidopteran insect of the Saturniidae family produces tasar silk (Vanya silk) of commercial importance [1]. The insect is having 44 ecoraces distributed along central India with varied phenotypic, physiological and behavioral characters. It is a polyphagous insect and feeds on *Terminalia tomentosa*, *Terminalia arjuna* etc.,

In insects, the growth and development is associated with protein metabolism [2]. In insects various biochemical changes occurs in the total concentration of haemolymph amino acids and proteins during metamorphosis [3]. In silkworms, the protein synthesis activity of the body wall and the midgut decreased when the larvae began to moult and increased from the midstage of the moulting period [4]. Recently, some work has been carried out on the haemocytes and protein changes in tasar silkworm [5]. The growth of silkworm is correlated with the synthesis of carbohydrates, proteins, amino acids and various enzymes [6]. During fifth instar the larvae spin a cocoon for protection. After this larva change into the pupa, finally adult moth emerges from cocoon. Various proteins are synthesized in the fat bodies and haemolymph of silkworm which are required to participate in physiological process. During insect metamorphosis, profound biochemical changes occur in the haemolymph,

particularly in the levels of proteins[7]. It is evident that profound changes in protein metabolism take place at various periods during insect development. Transformation from larva to pupa at the time of metamorphosis is accompanied by characteristic variations in the patterns of aminoacids, peptides, and proteins [8]. Physiological and biochemical studies on insect development have provided a reasonable picture of the causal relationships of protein synthesis, growth, and differentiation [9].

II. MATERIALS AND METHODS

The present work was performed by collecting Daba T.V cocoons as per the standard norms such as weight, colour, size of cocoons and length of the peduncle from the forest patches of Jakaram, Warangal District, during the third crop. The cocoons were preserved in the wire mesh cages of size 2 ft x2 ft x2 ft under temperature of 29 ± 1 °C and humidity 70 % \pm 1 %. The emerged moths were tested for pebrine disease [10]. Disease free layings were collected and incubated at 25- 30°C temperature and 70- 75% humidity and reared on Terminaliarjuna plantation till cocooning.

First day to sixth day 5th instar larvae, spinning day 1(SP1), spinning day 2(SP2) and pre pupa(PP) were used for experiment. Haemolymph was collected in the test tubes by cutting the proleg. 1mg of Thiourea was added to the test tubes to prevent melanisation. After centrifugation at 15000 rpm for 15min the supernatant was collected and stored at -20°C for further analysis. The fat body was isolated in cold condition by using Bodenstein's Ringer solution. 20% (W/V) homogenates of fat body was prepared in 50mM Tris -HCl Buffer (pH- 7.0) and centrifuged at 10,000 rpm for 20 min. Supernatant was collected for quantitative estimation of biochemical components. Total proteins and Aminoacids were estimated according to the standard procedures [11,12]. Enzyme like Acid phosphatase was estimated according to [13].

For haemocytes estimation the haemolymph was drawn into Thoma white blood pipette upto 0.5 mark and diluted upto 11 mark with tauberyeager fluid. The pipette was shaken for 5 minutes and first three drops were discarded. 0.5 ml of the haemocyte suspension was mixed with 0.5 ml of PBS and 0.5 ml of trypan blue. 0.2ml of this suspension is placed on hemocytometer and number of viable cells were counted under the microscope. The viable cell does not permit the entry of dye this distinguishes dead cells from living. Percentage of viable cells was calculated as

% of viable cells: =Number of viable cells / Number of viable cells + Number of dead cells X100

III. STATISTICAL ANALYSIS

Each assay was replicated 3 times. Values were expressed as mean \pm SE.

IV. RESULTS AND DISCUSSION

Table 1 shows protein concentration in haemolymph and fat body of fifth instar larvae and also prepupa. Total protein concentration varied in haemolymph and fatbody during day 1 to day 6 of 5th instar larvae and spinning day 1, day 2 and prepupal stage. Total protein concentration of both haemolymph and fat body increased up to day 6th day of fifth instar larvae from first day but a gradual reduction in protein content is observed during first day, second day of spinning and also in prepupal stage. During metamorphosis the protein concentration decreases gradually in haemolymph [14]. Different types of storage proteins are seen in silkworm which



involved in metamorphosis of silkworm. The total volume of protein increases exponentially with age in haemolymph of honey bees [15]. High transaminase activity results in high protein content [16]

Table 2 shows amino acid concentration in haemolymph and fat body of fifth instar larvae and also prepupa. Variation in amino acid concentration observed in haemolymph and fat body during day 1st to day 6th day of 5th instar larvae and spinning day 1, day 2 and prepupal stage. Total amino acid concentration of both haemolymph and fat body found decreased up to day 6th day of fifth instar larvae from first day but a gradual increase in amino acid content was observed during first day, second day of spinning and also in prepupal stage. Decrease in amino acids correlates with the increase in proteins. Decrease in the free amino acid content may indicate the possibility of active feeding of amino acid in Krebs's cycle and glycolytic pathway to meet the emergent energy needs as well as their utilization in the production of some new proteins synthesized to cope with the low temperature stress [17,18]. The decrease in amino acids in the fat body and silk gland could be their conversion to pyruvate by enzymes [19].

Acid phosphatase used as a marker enzyme for lysosomes and for apoptosis, involved in the degradation of insect tissues and thus involved in metamorphosis or transformation [20]. Present results show that acid phosphatases vary in the fat body of the silkworm during larval to pupal transformation. The activity of Acid phosphatase increased gradually till sixth day of fifth instar larvae followed by spinning day 1 and spinning day 2 and pre pupal stage (Fig 1).

Haemocytes are circulating cells and found in the haemolymph of insects responsible for the defense mechanism against foreign body [21]. Haemocytes perform various physiological functions in the insect body, changes in total haemocyte counts of particular insect directly or indirectly affect the insect [22]. The haemolymph is store house of various kinds of proteins and enzymes [23]. These proteins function as the main reservoirs for the supply of amino acids during larval moults and metamorphosis [24]. Results also show that haemocyte count varies from larval to pupal transformation. The total viable haemocytes count increased from first day to sixth day of fifth instar larvae whereas it got decreased in first day spinning, second day spinning and in prepupal stage (Fig.2).

V. CONCLUSION

Thus in conclusion protein levels, viable haemocytes are more in larvae but less during spinning and pupal stages. The amino acid levels are more in larvae and less in pupa whereas the activity of acid phosphatase is higher in pupa than in larva.

VI. ACKNOWLEDGEMENTS

The author would like to thank UGC-New Delhi for providing financial assistance in the form of Post-Doctoral Fellow.



REFERENCES

- [1] N.Suryanarayana and A.K. Srivastava, Monograph on Tropical Tasar Silkworm. Central Tasar Research and Training Institute, Ranchi, India, 2005, pp: 1-87.
- [2] A.K.Sinha, S.K.Chaudhury and K.Sen Gupta, Changes in free amino acids in the larval and pupal haemolymph of *Antheraea mylitta* Drury reared on *Terminalia arjuna* and *T. tomentosa*. Indian Journal of Sericulture, 1987, 27, 95-108.
- [3] B.A.Kilby, Intermediary metabolism and the insect fat body. Biochemical Society Symposia. 1965, 25:39-48.
- [4] M.J. Nagota, Silkworm developmental studies. Journal of Sericultural Science, 1976, 45, 328- 336.
- [5] D.D.Barsagade and D.B.Tembhare, Haemolymph amino acids and protein profile in the tropical tasar silkworm, *Antheraea mylitta* (Drury) (Lepidoptera: Saturniidae). Entomon. 2004, 29, 261-266.
- [6] V.C.Sivaprasad, C.Chandrasekharaiah, S.Misra, K.P.K.Kumar and Y.U.M.Rao, Screening of silkworm breeds for tolerance to *Bombyx mori* nuclear polyhedrosis virus (BmNPV), International Journal of Industrial Entomology, 2003, 7, 87-91.
- [7] Awauibsabhat, A.M. Sofi, M.A. Malik and Firdose Ahmad Malik. Studies on larval hemolymph protein levels of selected races of the silkworm, *Bombyx mori* L. under temperate climates of Kashmir. An international quarterly journal of life science. 2011; 6 (4): 533-536.
- [8] S.S.Rath, Raj Narain and B.C.Prasad, Impact of physiological condition of fifth instar larvae of *Antheraea mylitta* on rate of feeding and assimilation and its nutritional requirements. Proceedings of National Academy of Science, India, Section-B, Biology, 2004, 74, 237-243.
- [9] H.Ohta, T.Y.Utsumi and Y.Ozoe, Amino acid residues involved in the interaction with tyranine in *Bombyx mori* tyranine receptor. Insect Molecular Biology, 2004, 13, 531-538.
- [10] L. Pasteur, Etudes sur la maladie des vers a soie. Gauthier-Villars, Paris. Tome I, 1870, 322-327.
- [11] H.Lowry, N.L.Rosebrough, A.L.Far and R.J.Randal, Protein measurement with Folin reagent, Journal of Biology and Chemistry, 1951, 193, 265-275.
- [12] S.Moore and W.H. Stein, A modified ninhydrin reagent for the photometric determination of amino acids and related compounds, nitrogenous compound in urine of mature silkworm larvae, *Bombyx mori*, Applied entomology zoology, 1954, 15, 6065.
- [13] H. G. de Couet and A. D. Blest, The retinal acid phosphatase of a crab, *Leptograpsus*: Characterisation, and relation to the cyclical turnover of photoreceptor membrane. Journal of comparative physiology, 1982, 149(3), 353-362.
- [14] P.Sumithra, C.P.Britto and M.Krishnan, Modes of cell death in the pupal perivisceral fat body tissue of the silkworm *Bombyx mori* L. Cell and Tissue Research. 2010, 339(2):349-358.
- [15] W.T.Queenie Chan and J.Leonard Foster, Changes in protein expression during honey bee larval development, Genome biology, 2008, 156-162.
- [16] H.Watanabe and M.Kobayashi, Effect of virus infection on protein synthesis in silk gland of *Bombyx mori* L, Journal of invertebrate pathology, 1976, 14, 102-103.
- [17] Z.Zhao, Progress in the research on mechanism of insects cold-hardiness, 1997, 4(3):265-276.



[18] M.Forcella, E.Berra, R.Giacchini and P.Parenti, Antioxidant defenses preserve membrane transport activity in larvae exposed to anoxia, 2007, 65:181-194.

[19] Singh Anitha, K.Sharma Ratnesh, Sharma Bechan, Low temperature induced alterations in certain biochemical constituents of 5th instar larvae of *philosamiaricini* (Lepidoptera: Saturniidae), Insect Physiology, 2010, 3, 11-16.

[20] R.Halaby, M.L.Martinez, R.A.Lockshin and Z.Zakeri, 20-Hydroxyecdysone induces apoptosis in the labial gland of *Manduca sexta*. Journal of Research on the Lepidoptera, 2003, 37, 3-10.

[21] K. Rajitha, G. Savithri and P. Sujathamma, Haemocyte population dynamics in fifth instar silkworm *Bombyx mori* L inoculated with *Beauveria bassiana* (Bals.) vuill. International Journal of Agriculture Science Research, 2013, 3(2), 265-276.

[22] M.M. Essawy and I.A.I.Saad, Effect of potassium, sodium and its mixture supplementation on 1-blood volume, total and absolute haemocyte counts in mulberry silkworm *Bombyx mori* (L.) Journal of Entomology and Zoology Studies 2013, 1(6), 92-96.

[23] V.B.Wigglesworth, The principles of Insect physiology, 7th ed. Chapman and hall, London. 1972.

[24] M.Nagata and J.Kobayashi, Effects of nutrition on storage protein concentrations in the larval hemolymph of the silkworm *Bombyx mori*. Journal of Sericultural Science, 1990, 59(6), 469-474.

Table 1 Amino acid levels in haemolymph and Fat body during transformation from fifth instar larva to pupa of *Antheraea mylittadrury* (*Daba T.V*)

Source	1 st Day	2 nd day	3 rd day	4 th day	5 th day	6 th day	SP 1	SP2	PP
Haemolymph (mg/100ml)	998.6 ±3.5	989.4 ± 2.5	982.8 ± 3.6	975.6 ± 2	962.8 ± 3.8	942.4 ± 4	972.8 ± 6	985.6 ± 8	992.6 ± 5
Fat body (µg/100mg)	2.78 ± 0.18	2.63 ± 0.15	2.54 ± 0.14	2.49 ± 0.16	2.45 ± 0.12	2.38 ± 0.18	2.58 ± 0.15	2.64 ± 0.18	2.76 ± 0.12

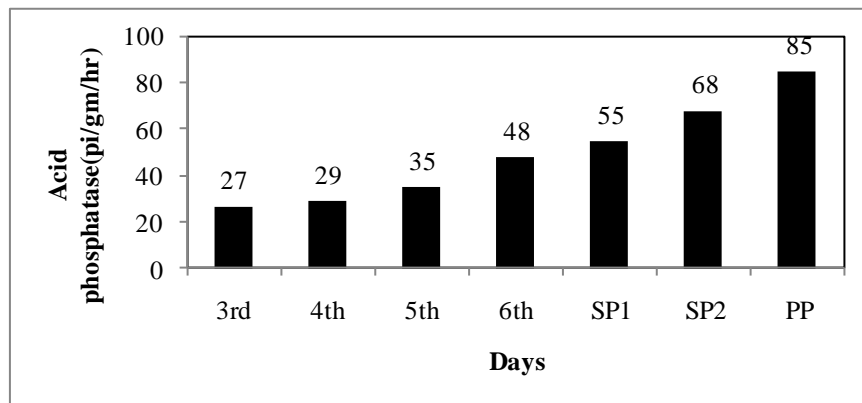
SP1-Spinning day 1, SP2-Spinning day 2, PP-Prepupa

Table 2 Protein levels in the haemolymph and fat body during transformation from fifth instar larva to pupa of *Antheraea mylittadrury* (*Daba T.V*)

Source	1 st day	2 nd day	3 rd day	4 th day	5 th day	6 th day	SP 1	SP2	PP
Haemolymph (mg/100ml)	18.5 ± 0.5	19.6 ± 0.6	25.6 ± 1.4	27.8 ± 0.5	32.5 ± 0.6	35.8 ± 0.8	16.6 ± 0.4	12.4 ± 0.8	10.6 ± 0.5
Fat body (mg/g)	78.5 ± 0.25	82.2 ± 1.15	86.2 ± 0.08	98.8 ± 1.12	108.8 ± 0.08	112.6 ± 2.14	48.4 ± 1.15	41.8 ± 1.16	38.6 ± 0.06

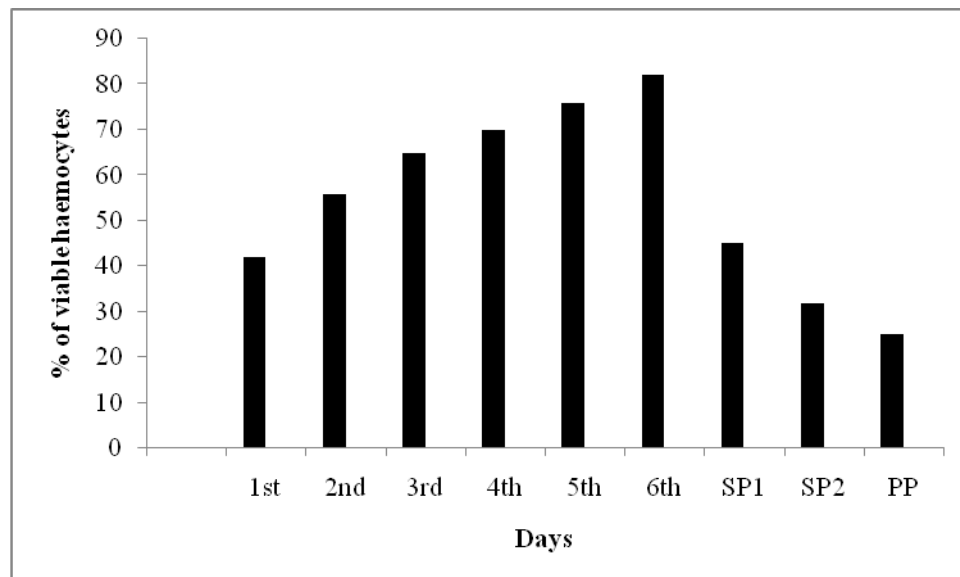
SP1-Spinning day 1, SP2-Spinning day 2, PP-Prepupa

Fig .1 Acid phosphatase activity in fat body during larva to pupa transformation



SP1-Spinning day 1, SP2-Spinning day 2, PP-Prepupa

Fig.2 Total viable haemocyte count during transformation of fifth instar larva to pupa in *Antheraea mylitta drury (Daba TV)*



SP1-Spinning day 1, SP2-Spinning day 2, PP-Prepupa